

## Laboratory Patient Collection Instructions Stool (LTR74542)

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Revision: 1.10

### Collecting Stool Samples for Bacterial PCR Screen, Clostridium difficile Toxin, Parasite PCR Screen, and/or Ova and Parasites

Procedure	Follow instructions below. If any questions, contact the Laboratory Information Centre at 403-770-3600.
Preparing	<b>DO NOT</b> use a laxative before collecting the stool sample, including mineral or castor oil.
Caution	<ul style="list-style-type: none"><li>The SAF solution is poisonous. Keep out of reach of children.</li><li>If swallowed, drink lots of milk or water and IMMEDIATELY call the Alberta Poison &amp; Drug Information Service (PADIS) at 1-800-332-1414.</li></ul>
Labelling the sample	<p><b>Label the collection container(s) with:</b></p> <ul style="list-style-type: none"><li>Laboratory- or clinic-issued patient label <b>or</b></li><li>Write the patient's <b>full first and last name</b>, and <b>Provincial Health Number (PHN)</b> or Government issued identification (Federal, Military, RCMP, Immigration).</li><li>Write <b>date and time of collection</b>.</li><li>For multiple samples to be collected on different days, also write #1, #2, etc. on each container to indicate the order the samples were collected.</li></ul>
Lab requisition	<ul style="list-style-type: none"><li>Ensure the laboratory requisition is labeled by the clinic with the correct patient information (matching that on specimen).</li><li>For Ova &amp; Parasite test requests, ensure <b>Stool Ova &amp; Parasite History Form</b> (Form – DMIC – 74535) is completed by your physician and is submitted with the Laboratory Requisition.</li><li><b>Print the date and time</b> the sample was collected on the lab requisition. Place the requisition in the <b>outside</b> pocket of the plastic resealable bag.</li><li>For <b>multiple samples</b>, write the date and a time the sample was collected on the requisition with the corresponding number belonging to the sample (e.g., #1, #2) in the "Date &amp; Time Collected:" space at the bottom of the Microbiology requisition for each sample collected.</li></ul> <p><b>Samples not labelled as required or those that do not have a completed requisition will NOT be tested.</b></p>
Before collecting	<ul style="list-style-type: none"><li>Obtain sterile collection container(s) from clinic or laboratory staff.</li><li>Depending on the tests ordered, you may receive an empty sterile container (usually with an orange lid), or a Stool Collection Kit which includes one empty sterile container (usually with an orange lid) and one container with a preservative called SAF (yellow lid). For all testing to be performed, stool must be added to <b>both</b> containers, if provided.</li><li>Please check expiration date on the yellow top container. If expired, get a new container from the laboratory.</li><li>If you need to collect more than one stool sample (e.g., three), collect each sample on a different day.</li></ul>

## Collecting the Sample

**Wash hands** with soap and water before collecting sample.

### Adults:

- Urinate (pee) completely first to prevent contamination of the sample with urine.
- Collect the stool sample into a clean, dry disposable container, waxed cardboard container, or on to plastic wrap placed under toilet seat.
- **DO NOT** let water touch the stool sample.

### Infants (diaper):

- **DO NOT** submit stool sample in diapers. For small children with diarrhea, or liquid stool, fasten a diaper liner (e.g., plastic kitchen wrap) to the diaper using childproof safety pins. Remove the stool sample from the diaper using a disposable plastic spoon and put into the container(s) following the steps below.

### Adding Stool to Collection Containers:

- Take parts of the stool that look bloody or slimy (contain mucous/pus).
- **Add stool to all containers that are provided:**

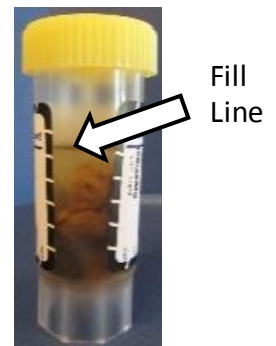
#### **Sterile Container (usually orange lid):**

- Using a wooden stick, fill the empty container at least 1/3 full.
- **Do not** fill more than half full.
- Replace lid. Ensure lid is tightly closed and not leaking.



#### **SAF Container (yellow lid):**

- Using wooden stick or fork/spoon provided with container, fill SAF fixative container by adding enough stool to bring the fluid **as close to the fill line as possible but not above.** This will require approximately 1 teaspoon (5 mL) of stool.
- **Do not overfill** as testing cannot be performed on overfilled SAF containers.
- Replace lid. Ensure lid is tightly closed and not leaking. Shake the stool and SAF until well-mixed.
- **If a whole worm is identified**, place in SAF container (yellow lid).



## Collecting the sample (continued)

Place sample container(s) in the provided plastic resealable bag(s). The requisition should be in the **outside** pocket. **Do not** put the requisition inside the bag with the sample container.

**Wash hands** thoroughly with soap and water after collecting the sample

## Delivering the sample

- **Transport the sample(s) and requisition(s) to the laboratory as soon as possible** (within 8 hours after collecting). If collecting multiple specimens on different days, deliver each specimen on the day collected. **Do not wait until all are collected.**
- If the stool sample cannot be delivered to the laboratory immediately after collection, keep refrigerated and deliver to laboratory within 24 hours.